

VeriKine[™] Mouse Interferon Alpha ELISA Kit Certificate of Analysis & Protocol

Assay Range: 12.5 - 400 pg/ml Compatibility: Tissue Culture Media Assay Length: 27 hr 15 min

Catalog No: 42120-1

Lot No: Expiration:

Store all components at 2-8°C

Kit Components	Part No.	Lot No.	Quantity
Plate(s)	SMP167		1
Plate Sealers	N/A	N/A	4
Wash Solution Concentrate	SMP022-60		50 ml
Mouse IFN-Alpha Standard, 10,000 pg/ml	SMP166-1		1 vial
Sample Buffer	SMP169-60		50 ml
Antibody Concentrate	SMP170-1		1 vial
Concentrate Diluent	SMP024-15		15 ml
HRP Conjugate Concentrate	SMP179-180		1 vial
Assay Diluent	ASD-15		15 ml
TMB Substrate Solution	KET-15		15 ml
Stop Solution	SCY-15		15 ml

Product Performance Specifications

	Sample Buffer	RPMI (10% FBS)*
Intra-Assay CV	≤ 8%	≤ 10%
Inter-Assay CV	≤ 10%	≤ 10%

Spike Recovery: 115% (range 82 – 135%)

 * We have noticed variability in the %CV between replicates of individual points on the standard curves prepared in MEM (10% FBS). The range of the variations observed is from 0.3 - 28%. These variations do not affect the product's ability to accurately measure Mouse IFN- α independently spiked in MEM (10% FBS).

Authorization

Released by: _		
Date:		

Visit PBL's website
(pblassaysci.com/documentation)
for additional information including
technical data sheets

Note: The quantitation of mouse interferon alpha in serum and plasma samples using this product has not been fully evaluated by PBL.

CAUTION: Wash Solution Concentrate, Sample Buffer and HRP/Ab Diluent contain 0.1% Kathon CG/ICP as a preservative; components should be handled with appropriate safety precautions and discarded properly. For further information, consult the safety data sheet (SDS).

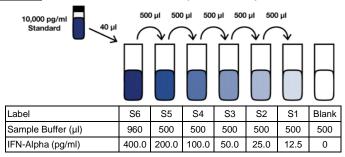
PREPARATION OF REAGENTS

<u>Wash Solution</u>: Wash Solution Concentrate may contain crystals; place in a warm water bath and gently mix until completely dissolved. Prepare a 1:20 working wash solution (e.g. Add 50 ml Wash Solution Concentrate to 1000 ml distilled or deionized water). Mix thoroughly before use. Keep at RT (22-25°C). (**Note:** Prepare fresh for each assay run.)

Mouse IFN-Alpha Standard Curve Preparation:

- a. Label six polypropylene tubes (S1 S6).
- b. Add volume of Sample Buffer or Sample Matrix to each tube as indicated in <u>Figure 1</u>. [Test samples may contain substances that can interfere with assay results, therefore it is recommended to run the IFN standard curve diluted in your Sample Matrix.]
- c. Using polypropylene tips, add 40 µl Standard to S6 and mix thoroughly. Change tips between each dilution.
- d. Transfer 500 μl of S6 to S5 and mix thoroughly. Repeat to complete series to S1. Set aside until use in step 1.

Figure 1: 6-Point Standard Curve Prepared in Sample Buffer



<u>Sample Preparation:</u> Thaw frozen sample tubes to Room Temperature (RT) (22-25 °C) in either tap water or between the fingertips. If samples require dilution, Sample Buffer is recommended. Keep at RT. Measurements in duplicate are recommended.

<u>Antibody Solution</u>: Dilute Antibody Concentrate in volume of Concentrate Diluent shown below. Keep at RT (22-25°C).

Micro-plate Strips Used	2	4	6	8	10	12
Antibody Concentrate (µI)						
Concentrate Diluent (ml)	2.0	3.0	4.0	5.0	6.0	7.0

HRP Solution: On Day 2, prepare within 15 minutes prior to use. Dilute HRP Conjugate Concentrate in volume of Assay Diluent shown below. Keep at RT (22-25°C).

Micro-plate Strips Used	2	4	6	8	10	12
HRP Conjugate Concentrate (µI)						
Assay Diluent (ml)	3.0	5.0	6.0	8.0	10.0	12.0

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ASSAY PROCEDURE

	Bring to RT (22-25°C)	Keep at 2-8°C		
Day 1	Wash Solution Concentrate	All other components		
Day 2	TMB Substrate Solution	All other components		
Da	Stop Solution			

- Incubations: Use plate sealers to cover the plate when directed. All incubations should be conducted in a closed chamber at RT (22-25°C)* keeping the plate away from drafts.
 - * Note: Transfer plate to 2-8°C for overnight incubation
- Plate Washing: All wells should be filled with a minimum of 300 µl of Wash Solution. Remove plate contents by inverting and blotting the plate on lint-free absorbent paper; tap the plate dry.

DAY 1

1. Determine the number of microplate strips required. We recommend running both the standard and samples at least in duplicate. Remove extra microtiter strips from the frame, seal in the foil bag provided and store at 2-8°C. Unused strips can be used in later assays.

Total well volume = 150 µl (Step A + Step B)

Step A: Add 100 µl of Standard, Test Samples or Blanks (Sample Buffer or appropriate dilution matrix) to each designated well.

Step B: Add 50 μ I of diluted Antibody Solution to every well. Change tips between each addition.

Cover with Plate Sealer and shake at 450 rpm at RT for 1 hour.

After 1 hour, transfer the plate to 2-8°C and incubate 20-24 hours. DO NOT SHAKE.

DAY 2

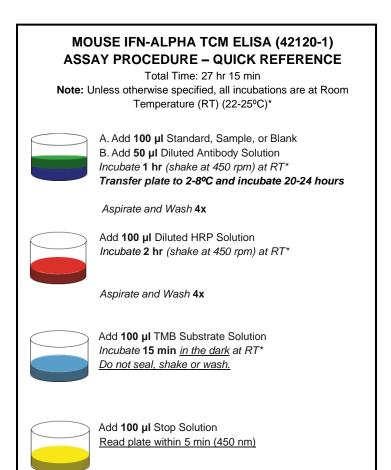
After 20-24 hours, empty plate contents and wash wells four times.

2. Add 100 µl of diluted HRP Solution to each well.

Cover with Plate Sealer and shake at 450 rpm at RT for 2 hours.

After 2 hours, empty plate contents and wash wells four times.

- 3. Add 100 μ l of TMB Substrate Solution to each well. Incubate in the dark at RT for 15 minutes. Do not use a Plate Sealer during the incubation. DO NOT SHAKE.
- **4.** After 15 minutes, DO NOT EMPTY THE WELLS AND DO NOT WASH. Add **100 \muI** of **Stop Solution** to each well.
- **5.** Using a microplate reader, determine the absorbance at 450 nm within 5 minutes after the addition of Stop Solution.



CALCULATION OF RESULTS

By plotting the optical densities (OD) using a 4-parameter fit for the standard curve, the interferon titer in the samples can be determined. A 4-parameter logistic plot with 1/y2 weighted analysis is recommended for obtaining optimal fit of standard curve OD values. Blank ODs may be subtracted from the standards and sample ODs to eliminate background.

Figure 2: Typical Standard Curve in Sample Diluent

