

# VeriKine<sup>™</sup> Cynomolgus/Rhesus IFN-Alpha Serum ELISA Kit Certificate of Analysis & Protocol

Assay Range: 25 - 1600 pg/ml Compatibility: Serum, Plasma, Tissue Culture Media Assay Length: 3 hr 15 min

Catalog No: 46100-1

Lot No: 7475

Expiration: August 31, 2022 Store all components at 2-8°C

Kit Components	Part No.	Lot No.	Quantity
Plate(s)	SMP135	K6771	1
Plate Sealers	N/A	N/A	4
Wash Solution Concentrate	SMP022-60	K6719	50 ml
Cyno/Rhesus IFN-Alpha 2 Standard, 10,000 pg/ml	SMP141-1	K6774	1 vial
Standard Diluent	SMP163-30	K6775	25 ml
Sample Buffer	SMP153-8	K6776	8 ml
Antibody Concentrate	SMP154-1	K6777	1 vial
HRP Conjugate Concentrate	SMP056-330	K6778	1 vial
Concentrate Diluent	SMP024-30	K6731	30 ml
TMB Substrate Solution	KET-15	210201D01	15 ml
Stop Solution	SCY-15	59172	15 ml

## **Product Performance Specifications**

	Standard Diluent	Cynomologus Monkey Serum	Tissue Culture Media (10% FBS)	
Intra-Assay CV	≤ 8%	≤ 8%	≤ 8%	
Inter-Assay CV	≤ 10%	≤ 15%	≤ 10%	

#### Authorization

Released by:

Date: December 13, 2021

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(pblassaysci.com/documentation)
for additional information including
technical data sheet

**CAUTION:** Sample Buffer, Standard Diluent, Wash Solution Concentrate and Concentrate Diluent contain 0.1% Kathon CG/ICP as a preservative and should be handled with appropriate safety precautions and discarded properly. For further information, consult the safety data sheet (SDS).

#### PREPARATION OF REAGENTS

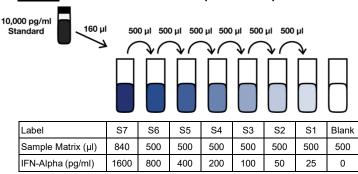
Wash Solution: Wash Solution Concentrate may contain crystals; place in a warm water bath and gently mix until completely dissolved. Prepare a 1:10 working wash solution (e.g. Add 50 ml Wash Solution Concentrate to 450 ml distilled or deionized water). Mix thoroughly before use. Keep at RT (22-25°C). (Note: Prepare fresh Wash Buffer for each assay run.)

## Cynomologus/Rhesus IFN-Alpha 2 Standard Curve Preparation:

**Note:** To avoid potential interference, it is recommended to run the standard curve diluted in endogenous IFN-free Sample Matrix. If Sample Matrix is unavailable, Standard Diluent may be used instead.

- a. Label seven polypropylene tubes (S1 S7).
- b. Add volume of Sample Matrix (e.g. Standard Diluent, Tissue Culture Media, Serum, Plasma) to each tube as indicated in <u>Figure 1</u>.
- c. Using polypropylene tips, add indicated amount of Cyno/Rhesus IFN-Alpha 2 to S7 and mix gently. Removed indicated amount from S7 and add to S6. Repeat to complete series to S1. Change tips between each dilution.

Figure 1: 7-Point Standard Curve Prepared in Sample Matrix



<u>Sample Preparation:</u> Thaw frozen sample tubes to Room Temperature (RT) (22-25°C) in either tap water or between the fingertips. If samples require dilution, prepare using Sample Matrix. Keep at RT until use. Measurements in duplicate are recommended.

Antibody Solution: 15 minutes prior to use in step 2, dilute Antibody Concentrate in the volume of Concentrate Diluent as shown below. Keep at RT (22-25°C).

Micro-plate Strips Used	2	4	6	8	10	12
Antibody Concentrate (µI)	20	40	60	80	100	120
Concentrate Diluent (ml)	2.0	4.0	6.0	8.0	10.0	12.0

<u>HRP Solution</u>: 15 minutes prior to use in step 3, dilute HRP Conjugate Concentrate in the volume of Concentrate Diluent as shown below. Keep at RT (22-25°C).

Micro-plate Strips Used	2	4	6	8	10	12
HRP Conjugate Concentrate (µI)	10	20	30	40	50	60
Concentrate Diluent (ml)	2.0	4.0	6.0	8.0	10.0	12.0

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#### **ASSAY PROCEDURE**

Bring to RT (22-25°C)	Keep at 2-8°C		
Plate/Plate Sealers	Cyno/Rhesus IFN-Alpha 2 Standard		
Wash Solution Concentrate	Antibody Concentrate		
Matrices/Samples	HRP Conjugate Concentrate		
Standard Diluent			
Sample Buffer			
Concentrate Diluent			
TMB Substrate Solution			
Stop Solution			

- Incubations: Use plate sealers to cover the plate when directed. All
  incubations should be conducted in a closed chamber at RT (2225°C), keeping the plate away from drafts.
- Plate Washing: All wells should be filled with a minimum of 250 µl of Wash Solution. Remove plate contents by inverting and blotting the plate on lint-free absorbent paper; tap the plate dry.
- 1. Determine the number of microplate strips required. We recommend running both the standard and samples at least in duplicate. Remove extra microtiter strips from the frame, seal in the foil bag provided and store at 2-8°C. Unused strips can be used in later assays.

Total well volume = 100 µl (Step 1 + Step 2)

Step 1: Add 50 µI of Sample Buffer to every well.

Step 2: Add 50  $\mu$ l of Standard, Test Samples or Blanks (Standard Diluent or appropriate dilution matrix) to each designated well.

Cover with Plate Sealer and shake at 450 rpm at RT for 1 hour.

After 1 hour, empty plate contents and wash wells two times.

2. Add 100 µI of diluted Antibody Solution to each well.

Cover with Plate Sealer and shake at 450 rpm at RT for 1 hour.

After 1 hour, empty plate contents and wash wells three times.

3. Add 100  $\mu l$  of diluted HRP Solution to each well.

Cover with Plate Sealer and shake at 450 rpm at RT for 1 hour.

After 1 hour, empty plate contents and wash wells four times.

- **4.** Add **100**  $\mu$ I of **TMB Substrate Solution** to each well. Incubate **in the dark** at RT for 15 minutes. Do not use a Plate Sealer during the incubation. DO NOT SHAKE.
- 5. After 15 minutes, DO NOT EMPTY THE WELLS AND DO NOT WASH. Add 100  $\mu l$  of Stop Solution to each well.
- **6.** Using a microplate reader, determine the absorbance at 450 nm within 5 minutes after the addition of Stop Solution.

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# CYNO/RHESUS IFN-ALPHA SERUM ELISA (46100) ASSAY PROCEDURE – QUICK REFERENCE

Total Time: 3 hr 15 min

Note: All incubations are at Room Temperature (RT) (22-25°C)\*



- 1. Add **50 μI** Sample Buffer
- 2. Add **50 µl** Standard, Sample or Blank *Incubate* **1 hr** (*shake at 450 rpm*) *at RT\**

Aspirate and Wash 2x



Add **100 μI** diluted Antibody Solution Incubate **1 hr** (shake at 450 rpm) at RT\*

Aspirate and Wash 3x



Add **100 μI** diluted HRP Solution *Incubate* **1 hr** (shake at 450 rpm) at RT\*

Aspirate and Wash 4x



Add **100 µl** TMB Substrate Incubate **15 min** in the dark Do not seal, shake or wash.



Add **100 μl** Stop Solution Read plate within 5 min (450 nm)

### **CALCULATION OF RESULTS**

By plotting the optical densities (OD) using a 4-parameter fit for the standard curve, the interferon titer in the samples can be determined. Blank ODs may be subtracted from the standards and sample ODs to eliminate background. The conversion factor of about 3-5 pg/unit is applicable for cynomologus/rhesus IFN-Alpha 2 where units are determined by comparison to human IFN-Alpha 2 international standard. Nevertheless, this conversion factor is only an approximation.

Note: The lowest limit of quantitation (LLOQ) is 25 pg/ml. Concentrations of unknown samples that measure < 25 pg/ml are suspect.

Figure 2: Typical Standard Curve

